

Molecular vibration-sensing component in *Drosophila melanogaster* olfaction

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A common explanation of molecular recognition by the olfactory system posits that receptors recognize the structure or shape of the odorant molecule. We performed a rigorous test of shape recognition by replacing hydrogen with deuterium in odorants and asking whether *Drosophila melanogaster* can distinguish these identically shaped isotopes. We report that flies not only differentiate between isotopic odorants, but can be conditioned to selectively avoid the common or the deuterated isotope. Furthermore, flies trained to discriminate against the normal or deuterated isotopes of a compound, selectively avoid the corresponding isotope of a different odorant. Finally, flies trained to avoid a deuterated compound exhibit selective aversion to an unrelated molecule with a vibrational mode in the energy range of the carbon–deuterium stretch. These findings are inconsistent with a shape-only model for smell, and instead support the existence of a molecular vibration-sensing component to olfactory reception.

Olfactory systems perform remarkable feats of molecular recognition, but although much is known about the neurophysiology of olfaction (1–5), how olfactory receptors "read" molecular structure remains unknown. Parts of odorant molecules (odotopes) have been proposed to engage particular receptors in a "lock-and-key" manner and this molecular shape recognition mechanism is thought sufficient for odor discrimination (2). An alternative hypothesis (6) posits that molecular vibrations of all atoms, or of particular functional groups of odorant molecules, contribute to odor recognition, and odorants with similar vibrational spectra should elicit similar olfactory responses (7). Molecules in which deuterium replaces nonexchangeable hydrogens constitute appropriate probes to test these alternatives because deuteration does not alter atom size or bond length or stiffness (8). Thus, the conformation of a deuterated molecule should be identical to that of a hydrogen-only (i.e., normal) odorant and, according to the molecular recognition theory, the two isotopes should smell identical. However, atoms in a molecule vibrate in normal modes at particular energies that depend on the molecular structure. The doubling of nuclear mass upon deuteration will change all the vibrational modes of an odorant molecule to differing extents. The second hypothesis predicts that deuteration will alter its smell in comparison with the hydrogen-only isotope. A recent attempt to test the latter hypothesis in humans was rather inconclusive, potentially confounded by interference from previous experience, olfactory training or habituation of the subjects, or shortcomings of human olfaction as suggested by the authors (9). A major experimental difficulty is that, in general, isotopes are prepared and purified differently, and therefore perceived differences in smell could be attributed to different impurities. Purity can, in principle, be ensured by smelling single-molecule peaks exiting a gas chromatographic column, but this makes simultaneous two-compound comparisons difficult and animal experiments harder still.

Here we describe a different approach to test whether molecular vibrations contribute to recognition of odor character. We assumed that, if molecular vibrations are indeed detected, the deuterated isotopes of different odorants may share a common odor character, as deuteration shifts particular peaks—e.g., the carbon–deuterium (C–H) stretch—of the compound-specific vibrational spectra to lower frequencies. If so, the deuterium

odor character could be distinct and identifiable, irrespective of the structure and chemical properties of the odorant molecules that carry it. Significantly, we used *Drosophila* as unbiased and objective subjects to address this issue. They possess a relatively well understood olfactory system (10–13), exhibit keen olfactory discrimination (14–16), and can be conditioned to selectively avoid or seek odors with the use of established methodology (17, 18). We ask whether *Drosophila* can detect deuterium as a distinguishing molecular feature in odorant isotopes and a salient cue for conditioning. The results of these experiments provide support for the notion that flies can smell molecular vibrations.

Results

Spontaneous Differential Responses to Deuterated Odorants. Although deuteration does not appreciably change molecular shape, atom size, or bond length or stiffness, it doubles hydrogen mass, thus affecting the overall vibrational modes of an odorant. Therefore, if recognition of molecular shape alone was the sole determinant for odor character (2, 3), then flies should not respond differentially to deuterated [d, where $d_{(x)}$ denotes replacement of x nonexchangeable hydrogens with deuterium atoms] and nondeuterated/normal (i.e., H-) odorants. To address this hypothesis, we took advantage of the commercial availability of acetophenone (ACP) carrying three, five, or eight deuterium atoms (d_3 , d_5 , and d_8) in place of the respective hydrogens in the normal molecule (h-ACP). Equal amounts (75 μ L) of each odorant were diluted to 1 mL in isopropyl myristate and we quantified (Fig. 1A) the response of groups of flies to each odorant versus unscented air traversing the arms of a standard T-maze (*Materials and Methods*) (19, 20). When given a choice between normal ACP (i.e., h-ACP) and air, the excess flies present in the odor-delivering maze arm indicated that, at the dilution used, the odorant was attractive. However, replacing only three hydrogen atoms with deuterium eliminated this spontaneous preference as the flies distributed nearly equally in the arms of the maze. The d_5 -ACP was mildly aversive and the fully substituted d_8 -ACP was the most aversive of the four isotopes. Moreover, flies consistently avoided d_3 , d_5 , and d_8 ACP against equal concentration of h-ACP (Fig. S1). Therefore, a spontaneous switch in osmotaxis (14) to ACP was precipitated by substituting hydrogen atoms with deuterium.

To verify that flies are generally capable of spontaneous discrimination between airborne deuterated odorants and their normal counterparts, we additionally presented them with equal concentrations of normal and deuterated 1-octanol, normal and d_5 -benzaldehyde, or a normal–perdeuterated ACP pair. In confirmation of the previous results, h-ACP was preferred versus an equal concentration of the d_8 odorant (Fig. 1B). Reducing the concentration of the deuterated compound by 50% resulted in

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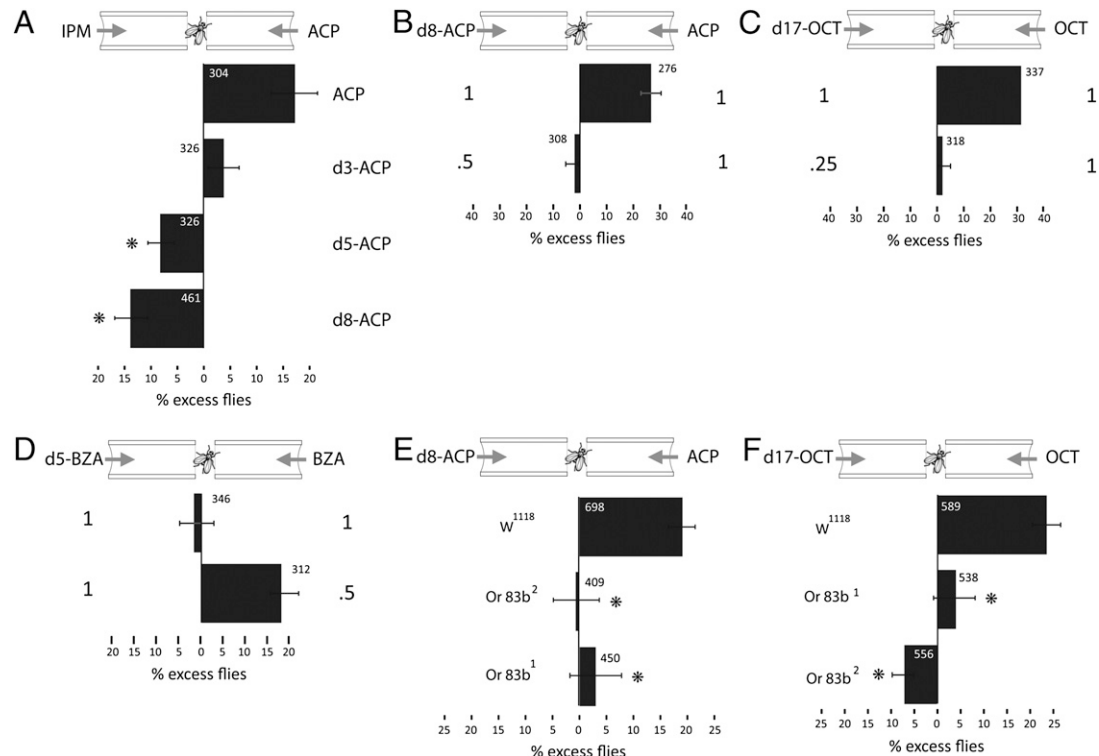
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Fig. 1. Differential spontaneous responses to odorants containing deuterium. The mean relative distribution of flies in the arms of the maze (% excess flies) carrying the indicated odorants $\pm$ SEM is shown in all graphs. This metric reflects the prevailing distribution within the arms of the maze of groups of 40 to 60 flies tested each time. The total number of flies tested in each group is shown and the number of groups tested is $n \geq 6$ for all groups. (A) Spontaneous responses to 75 μ L normal or d₃-, d₅-, or d₈-ACP, each diluted with isopropyl myristate (IPM) to 1 mL. Flies spontaneously preferred h-ACP over the solvent. In contrast, incorporation of five or eight deuterium atoms results in significantly different distribution ($P < 0.001$) from that toward h-ACP (attraction) to aversion. In



contrast, the response to d₃-ACP was not significantly different ($P = 0.012$) from that of h-ACP. (B) Flies discriminate against d₈-ACP if presented with equal amount (1:1) of h-ACP (75 μ L odorant/925 μ L IPM). However, a 50% reduction in the amount of deuterated odorant yielded an equal distribution of the flies in the arms (% excess flies not significantly different from zero), defined as a balanced maze. (C) Similarly equal amounts (200 μ L) of h-1-octanol (OCT) and d₁₇-1-octanol yielded strong discrimination against the deuterated odorant, which was eliminated upon reducing it by 75% (1:0.25). (D) In contrast, equal amounts of normal and deuterated benzaldehyde (90 μ L) did not result in differential discrimination. In accord, decreasing the amount of h-BZA resulted in differential avoidance of d₅-BZA. (E) The preferential discrimination against d₈-ACP was eliminated in *Or83b*¹ and *Or83b*² mutants ($P < 0.002$ and $P < 0.001$, respectively, Dunnett test). (F) Similarly, discrimination against d₁₇-octanol was eliminated in *Or83b*¹ and *Or83b*² mutants ($P < 0.001$ for both, Dunnett test).

balanced distribution of the flies in the maze arms, indicative of equal response to the two isotopes (i.e., balanced maze). Similarly, d₁₇-1-octanol was preferentially discriminated against versus equal concentration of the normal odorant. In this case, however, the deuterated isotope appeared even more aversive, as 75% reduction in its concentration equalized aversion with h-1-octanol (Fig. 1C). In contrast, flies did not exhibit spontaneous differential avoidance of d₅-benzaldehyde, which occurred only upon reducing the concentration of the normal odorant (Fig. 1D). These differences cannot result from reduced evaporation of the slightly heavier deuterated odorants because two of the three elicited increased aversion, known to be proportional to the concentration of airborne odorants in this T-maze system (14, 20, 21).

If the differential response to the deuterated odorants relied solely on olfaction and not any other sensory modality, it should be completely eliminated in anosmic mutants. To that end, we obtained two anosmic *Drosophila* strains carrying the null alleles *Or83b*¹ and *Or83b*² of the gene encoding the common subunit of the dimeric *Drosophila* olfactory receptor (13, 22, 23). Clearly, the spontaneous avoidance of deuterated d₈-ACP and d₁₇-1-octanol were eliminated and the mutant flies distributed equally in the maze arms as expected (Fig. 1E and F). Therefore, *Drosophila* use olfaction alone to discriminate between deuterated and normal odorants. Furthermore, the spontaneous discrimination against deuterated odorants indicates that they likely present to the flies recognizable, salient features distinct from their hydrogen counterparts.

Conditioned Discrimination of Isotopes. If the deuterated and normal isotopes of a pair exhibit salient odor character differences, then *Drosophila* should be able to associate either odorant with a punishing stimulus (20). To eliminate bias, the amounts of each odorant in a pair were adjusted as shown in Fig. 1B–D to yield

spontaneous preference as near zero as possible (i.e., balanced maze), so any postconditioning distribution changes would be a consequence of training. Flies were conditioned to associate electric foot-shock punishment with the presence of the deuterated or normal odorant of a pair. *Drosophila* successfully associated either odorant with punishment, demonstrated by the conditioned selective aversion of the shock-associated odor upon testing (Fig. 2). Flies shocked in the presence of the normal odorant distributed preferentially in the arm carrying its deuterated counterpart and vice versa. The shock-associated learning extended to h-benzaldehyde and d₅-benzaldehyde, for which the flies exhibited no spontaneous preference, suggesting, as expected (24), that conditioned discrimination is independent from spontaneous preference.

This was not an exclusive property of the w¹¹¹⁸ control strain (14), as experiments were repeated with Canton-S, a different WT strain, with identical results (Fig. S2A and B). We also reversed the order of stimulus presentation, such that the shock-associated odor was delivered immediately before testing. Flies continued to selectively avoid the shock-associated odor (Fig. 2C), eliminating the possibility that they were simply attracted to the odor presented last in the absence of the shock reinforcer. Thus, deuterated and normal odorants present salient differences to the fly olfactory system, which can be used to predict punishment or its absence by association with either isotope.

Generalization of Conditioned Isotope Discrimination Across Odorant Pairs. The results so far are consistent with an unambiguous difference between deuterated and normal odorants for all three pairs tested. Is the difference associated with the majority ($\geq 99\%$) compound or with impurities? Although the odorants used were of the highest purity available (Figs. S3 and S4), they

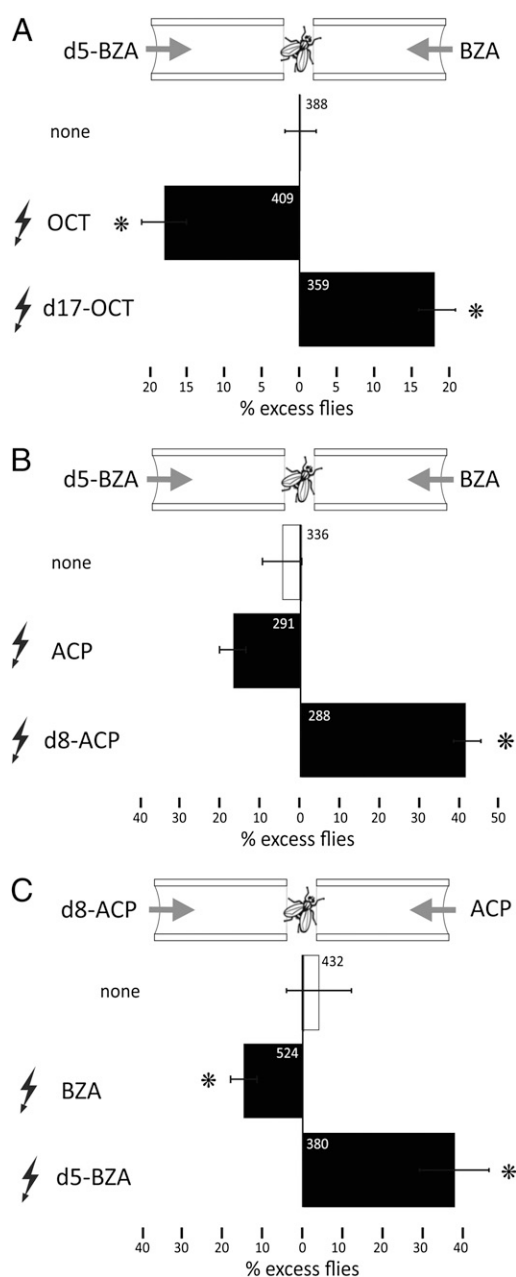
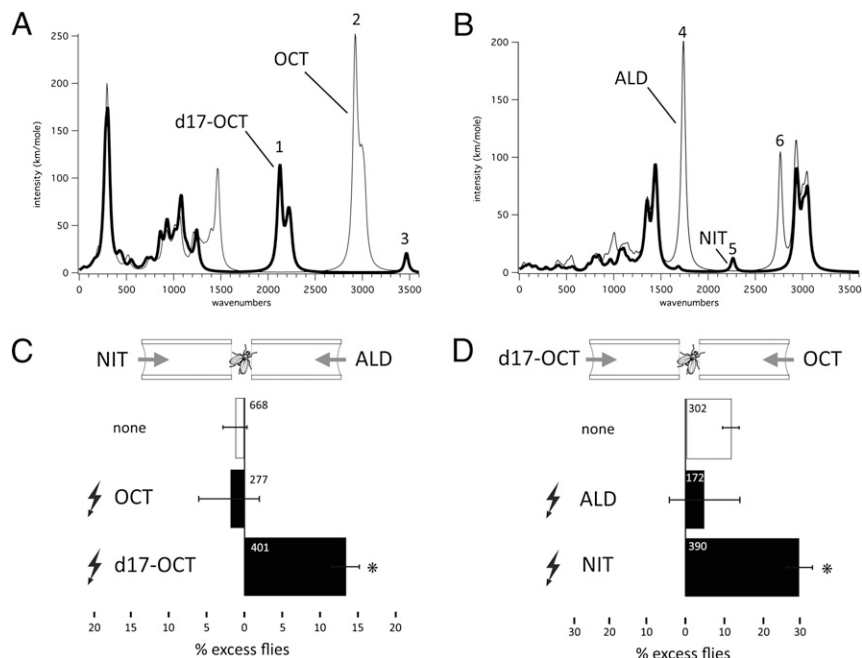


Fig. 4. *Drosophila* can be conditioned to selectively avoid a vibrational frequency. (A) Computed vibrational spectra (IR intensities) of h-OCT versus d₁₇-OCT. Salient spectral peaks are indicated on the graphs and show that deuteration of octanol shifts the group of C-H stretch vibrations from around 3,000 cm⁻¹ to 2,150 cm⁻¹. Deuteration also shifts downward all of the peaks on the fingerprint region (1,000–1,500 cm⁻¹). (B) Computed IR intensities of citronellal versus citronellyl nitrile with the salient spectral peaks indicated. The spectra of citronellal and citronellyl nitrile are remarkably similar in the fingerprint region. They differ chiefly in the vibrations involving the terminal functional group: in citronellyl nitrile the aldehyde carbonyl stretch around 1,750 cm⁻¹ is absent, replaced by a nitrile stretch around 2,150 cm⁻¹. The low-lying aldehyde C-H stretch vibration is also absent. The vibration band centered at 2,150 cm⁻¹ is the only one common to d₁₇-1-octanol and citronellyl nitrile but not present in h-octanol or citronellal. The spectra were computed using the Amsterdam Density Functional software at DZP/PBE level of theory. (C) *Drosophila* selectively avoid the molecular vibrations of deuterium. Flies conditioned to selectively avoid d₁₇-1-octanol exhibited strong preferential avoidance of citronellyl nitrile ($P < 0.001$ vs. naive), but flies punished to h-octanol did not selectively avoid citronellal ($P = 0.691$ vs. naive). The only common element potentially recognizable in the test odor pair to aid in selective avoidance is the overlap in the vibrational spectrum of the C-D bonds in d₁₇-1-octanol and the C≡N triple bond in citronellyl nitrile as illustrated in A and B. In contrast, they were not selective toward a novel odor without any recognizable molecular features. (D) In the converse experiment, flies conditioned to selectively avoid citronellyl nitrile exhibited highly significant avoidance of d₁₇-1-octanol as a testing odor ($P < 0.001$ vs. naive), but flies punished to citronellal did not selectively avoid h-octanol ($P = 0.999$ vs. naive).



differences because of the reduced evaporation of the heavier deuterated molecules. However, such differences would have to be greater than 30% (28), which is rather improbable under our experimental conditions, and flies seem to generalize concentration differences smaller than that (29). This explanation is additionally improbable because, except for benzaldehyde, the “heavier” deuterated compounds are more aversive than their normal isotopes. Moreover, this hypothesis is inconsistent with the identification of deuteration as a salient mediator of conditioned responses across chemically distinct odors and with the selective aversion of molecules with vibrational resonance in the range of the C-D stretch. Therefore, a parsimonious explanation for our results is strongly indicative of a molecular vibration-sensing component in *Drosophila* olfaction. This is also consistent with independent recent evidence suggesting contributions of the vibrational spectra of odorants in the electrophysiological response of isolated *Drosophila* olfactory receptors (31).

In the past there have been four main objections to the notion that olfaction detects molecular vibrations. First, that molecular shape is an adequate predictor of smell (2, 30, 32), which currently seems unlikely (33, 34). In fact, *Drosophila* has only 62 olfactory receptors (35), suggesting that a single receptor must generally respond to multiple odorants, but in addition that a single odorant can activate multiple receptors (10, 15). Clearly, therefore, the structural recognition mechanism alone does not suffice to explain odorant recognition, suggesting that additional properties of these volatile molecules likely contribute to the process. The second objection was to the main assertion of vibrational theory that odorants with similar spectra should produce similar olfactory responses and, conversely, molecules of identical molecular shape but distinct spectra should smell different. Our data are consistent with this hypothesis, at least in flies, and suggest that with gas chromatography-pure odorants, and perhaps with the use of aversive conditioning, similar effects may be revealed for vertebrates—even humans—as shown for perceptual discrimination of enantiomers (36). The third objection has been that no known biological mechanism could behave as the equivalent of a vibrational spectroscopy. However, the proposal (6) that olfaction uses inelastic electron tunneling

spectroscopy (IETS) has made the idea physically plausible. IETS is a quantum mechanism whereby electrons move from a donor to an acceptor site at constant total energy, although the acceptor is energetically lower than the donor (37). To satisfy conservation of energy, tunneling occurs only if a molecule is present between donor and acceptor, possessing vibrational mode(s) at or near this excess energy, absorbing it, and becoming excited. A modified IETS mechanism appropriate to proteins was recently described (26), suggesting a testable model applicable to olfactory receptors. Finally, it was objected that enantiomers with identical vibrations should always smell the same, whereas some smell different (38). Given that proteins are chiral, a shape-only theory cannot account for the identical odors of most enantiomer pairs. In contrast, this objection is easily answered by IETS because it exhibits the pronounced polarization effects expected when sensing molecules in fixed orientations such as enantiomers (6).

Individual olfactory receptors in *Drosophila* and other species can mediate excitatory and inhibitory responses to different odorants (10). Our data suggest that the vibrational mode and frequency of particular atoms and active groups of odorant molecules may also provide discriminatory cues that could broaden the recognition repertoire of odorant receptors, but still retain specificity. Currently, we do not know whether the same or different olfactory receptors are recruited to sense the normal and deuterated versions of the odorant molecules, and this question is currently under investigation broadly guided by the following considerations. A single receptor may recognize, by broad odotope features, a given odorant whose particular vibrational resonance may contribute to the odor-specific activity patterns of odorant receptor neurons (39), potentially modifying its particular odor character. This hypothesis predicts that receptors with distinct shape selectivities must also recognize similar vibrational modes and frequencies to explain our data of selective avoidance of citronellyl nitrile after conditioning with d₁₇-1-octanol and vice versa. By analogy to color vision, it is possible that the multiple olfactory receptors may be divided into spectral classes, the members of each class sensing the same vibrational range and differing in their affinity for molecules of different shapes and physical properties.

For example, we would expect to find several receptors in the 2,150-cm⁻¹ band, able to sense deuterated molecules but not their undeuterated counterparts.

If the proposed vibrational sensing component is arranged in broad frequency bands of 100 to 200 cm⁻¹ wide, 10 to 20 receptors would suffice to cover the entire vibrational range. However, why then do flies (and mammals) have many more receptors (2, 3)? A possible answer may be that the olfactory system has evolved to reconcile conflicting requirements, namely, to sense a wide range of odorants, thus being nonspecific, but also to possess high affinity to detect them in small concentrations. Therefore, multiple receptors may serve to ensure that one or more will always bind with adequate affinity to any molecule to provide broad olfactory recognition through the combined activities of multiple receptors. Testing these and other predictions is well within reach of available behavioral, imaging, and genetic techniques.

Materials and Methods

Drosophila were cultured as described previously (40) under a 12-h dark, 12-h light cycle. Mixed-sex groups of 2- to 3-d-old flies were lightly anesthetized and segregated in groups of 50 to 70 animals in vials containing food. Twenty-four hours later they were changed to fresh vials and placed in the dark to adapt 2 to 3 h before behavioral experiments commenced. Details of the behavioral procedures can be found in *SI Materials and Methods*.

All behavioral experiments were performed at 25 °C and 80% to 85% relative humidity. Training and testing were performed in complete darkness, with dim red light used only during manipulations not requiring exposure to odors. The standard T-maze was modified by replacing the odorant holding cups (20) with glass cylinders of 2.25 cm diameter and 14 cm height as described previously (19, 40). An air stream of 600 mL/min passing

over the meniscus carried the odors to the maze arms via silicon rubber tubing. The meniscus area was kept constant by maintaining the volume of the odorant and the solvent, isopropyl myristate (Fluka), at a total of 1 mL in all experiments. Deuterated compounds were from CDN Isotopes. The amounts of odorants added to isopropyl myristate to yield equivalent avoidances were 200 μ L of 1-octanol (Fluka) and 50 μ L of the fully deuterated d₁₇-1-octanol, 90 μ L of benzaldehyde (Fluka) and 90 μ L of perdeuterated benzaldehyde-d₅, 150 μ L of ACP (Fluka) and 75 μ L of the perdeuterated d₈-ACP, 100 μ L of citronellal and 100 μ L of citronellyl nitrile (gift of International Flavors and Fragrances). As for d₈-ACP (C6D₅COCD₃), 75 μ L d₃-ACP (C6H₅OCD₃), d₅-ACP (C6D₅OCH₃), and normal odorant were also used for discrimination experiments. Different sets of silicone rubber tubing holding cylinders and maze arms were used for each odor. Complete descriptions of the behavioral procedures are presented in *SI Materials and Methods*.

Computation of IR Intensity Spectra. Spectra were computed by using the Amsterdam Density Functional software package (www.scm.com) at DZP/PBE level of theory. Frequencies were computed numerically by differentiation of energy gradients in slightly displaced geometries. The force constants and hence the frequencies were computed by comparison of the gradients (in the harmonic approximation of the energy surface). Under these conditions, intensities are proportional to the change in dipole occurring during atom movements in a given vibrational mode (41). Accuracy of mode calculations is typically fewer than 10 wave numbers for molecules comprising C, O, N, and H atoms.

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1. Araneda RC, Kini AD, Firestein S (2000) The molecular receptive range of an odorant receptor. *Nat Neurosci* 3:1248–1255.
2. Axel R (2005) Scents and sensibility: A molecular logic of olfactory perception (Nobel lecture). *Angew Chem Int Ed Engl* 44:6110–6127.
3. Buck L, Axel R (1991) A novel multigene family may encode odorant receptors: a molecular basis for odor recognition. *Cell* 65:175–187.
4. Firestein S (2001) How the olfactory system makes sense of scents. *Nature* 413:211–218.
5. Malnic B, Hirono J, Sato T, Buck LB (1999) Combinatorial receptor codes for odors. *Cell* 96:713–723.
6. Turin L (1996) A spectroscopic mechanism for primary olfactory reception. *Chem Senses* 21:773–791.
7. Turin L (2002) A method for the calculation of odor character from molecular structure. *J Theor Biol* 216:367–385.
8. Wade D (1999) Deuterium isotope effects on noncovalent interactions between molecules. *Chem Biol Interact* 117:191–217.
9. Keller A, Vosshall LB (2004) A psychophysical test of the vibration theory of olfaction. *Nat Neurosci* 7:337–338.
10. Hallem EA, Ho MG, Carlson JR (2004) The molecular basis of odor coding in the *Drosophila* antenna. *Cell* 117:965–979.
11. Laissue PP, Vosshall LB (2008) The olfactory sensory map in *Drosophila*. *Adv Exp Med Biol* 628:102–114.
12. Nakagawa T, Vosshall LB (2009) Controversy and consensus: noncanonical signaling mechanisms in the insect olfactory system. *Curr Opin Neurobiol* 19:284–292.
13. Vosshall LB, Stocker RF (2007) Molecular architecture of smell and taste in *Drosophila*. *Annu Rev Neurosci* 30:505–533.
14. Acevedo SF, Froudarakis EI, Tsirova AA, Skoulakis EM (2007) Distinct neuronal circuits mediate experience-dependent, non-associative osmotactic responses in *Drosophila*. *Mol Cell Neurosci* 34:378–389.
15. Hallem EA, Carlson JR (2006) Coding of odors by a receptor repertoire. *Cell* 125:143–160.
16. Fiala A (2007) Olfaction and olfactory learning in *Drosophila*: Recent progress. *Curr Opin Neurobiol* 17:720–726.
17. Berry J, Krause WC, Davis RL (2008) Olfactory memory traces in *Drosophila*. *Progress in Brain Research: The Essence of Memory*, eds Sossin WS, Lacaille J-C, Castellucci VF, Bellville S (Elsevier, Amsterdam), Vol 169, pp 293–304.
18. Skoulakis EMC, Grammenoudi S (2006) Dunces and da Vincis: The genetics of learning and memory in *Drosophila*. *Cell Mol Life Sci* 63:975–988.
19. Pavlopoulos E, Anezaki M, Skoulakis EMC (2008) Neuralized is expressed in the α/β lobes of adult *Drosophila* mushroom bodies and facilitates olfactory long-term memory formation. *Proc Natl Acad Sci USA* 105:14674–14679.
20. Tully T, Quinn WG (1985) Classical conditioning and retention in normal and mutant *Drosophila melanogaster*. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 157:263–277.
21. Skoulakis EM, Davis RL (1996) Olfactory learning deficits in mutants for leonardo, a *Drosophila* gene encoding a 14-3-3 protein. *Neuron* 17:931–944.
22. Benton R, Sachse S, Michnick SW, Vosshall LB (2006) Atypical membrane topology and heteromeric function of *Drosophila* odorant receptors in vivo. *PLoS Biol* 4:e20.
23. Wicher D, et al. (2008) *Drosophila* odorant receptors are both ligand-gated and cyclic-nucleotide-activated cation channels. *Nature* 452:1007–1011.
24. Rescorla RA, Wagner AR, eds (1972) Black AH, Prokasy WK, eds (1972) A theory of Pavlovian conditioning: Variations in the effectiveness of reinforcement and nonreinforcement. *Classical Conditioning II: Current Research and Theory*, eds Rescorla RA, Wagner AR (Appleton-Century-Crofts, New York), pp 64–99.
25. Rescorla RA (1976) Stimulus generalization: Some predictions from a model of Pavlovian conditioning. *J Exp Psychol Anim Behav Process* 2:88–96.
26. Brookes JC, Hartoutsiou F, Horsfield AP, Stoneham AM (2007) Could humans recognize odor by phonon assisted tunneling? *Phys Rev Lett* 98:038101.
27. Bedoukian PZ, ed (1986) *Perfumery and Flavoring Synthetics* (Allured, Carol Stream, IL).
28. Borst A (1983) Computation of olfactory signal in *Drosophila melanogaster*. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 152:373–383.
29. Masek P, Heisenberg M (2008) Distinct memories of odor intensity and quality in *Drosophila*. *Proc Natl Acad Sci USA* 105:15985–15990.
30. Buck LB (2004) Olfactory receptors and odor coding in mammals. *Nutr Rev* 62 (suppl): S184–S188.
31. Guo S, Kim J (2010) Dissecting the molecular mechanism of *Drosophila* odorant receptors through activity modelling and comparative analysis. *Proteins* 78:381–399.
32. Rossiter KJ (1996) Structureminus signodor relationships. *Chem Rev* 96:3201–3240.
33. Sell CS (2006) On the unpredictability of odor. *Angew Chem Int Ed Engl* 45:6254–6261.
34. Turin L, Yoshii F (2003) Structure-odor relations: A modern perspective. *Handbook of Olfaction and Gustation*, ed Doty R (Marcel Dekker, New York).
35. Robertson HM, Warr CG, Carlson JR (2003) Molecular evolution of the insect chemoreceptor gene superfamily in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* 100 (suppl 2):14537–14542.
36. Li W, Howard JD, Parrish TB, Gottfried JA (2008) Aversive learning enhances perceptual and cortical discrimination of indiscriminable odor cues. *Science* 319:1842–1845.
37. Lambe J, Jaklevic RC (1968) Molecular vibration spectra by inelastic electron tunneling. *Phys Rev* 165:821–832.
38. Bentley R (2006) The nose as a stereochemist. Enantiomers and odor. *Chem Rev* 106: 4099–4112.
39. Wilson RI (2008) Neural and behavioral mechanisms of olfactory perception. *Curr Opin Neurobiol* 18:408–412.
40. Moresis A, Friedrich AR, Pavlopoulos E, Davis RL, Skoulakis EMC (2009) A dual role for the adaptor protein DRK in *Drosophila* olfactory learning and memory. *J Neurosci* 29: 2611–2625.
41. Fan L, Ziegler T (1992) Application of density functional theory to infrared absorption intensity calculations on main group molecules. *J Chem Phys* 96:6937–6941.